



International Journal of Applied Dental Sciences

ISSN Print: 2394-7489
ISSN Online: 2394-7497
IJADS 2022; 8(4): 169-172
© 2022 IJADS
www.oraljournal.com
Received: 05-09-2022
Accepted: 01-11-2022

Adrián Marcelo Villarreal García
Master's in Sciences Student,
Universidad Autonoma de Nuevo
Leon, Facultad de Odontología,
Monterrey, Nuevo Leon, 64460
ZIP, Mexico

Fanny Lopez Martinez
Professor, Universidad Autonoma
de Nuevo Leon, Facultad de
Odontología, Monterrey, Nuevo
Leon, 64460 ZIP, Mexico

Laura Roesch Ramos
Professor, Universidad
Veracruzana, Facultad de
Odontología, Veracruz, Veracruz,
Mexico

Evelyn Guadalupe Torres Capetillo
Professor, Universidad
Veracruzana, Facultad de
Odontología, Veracruz, Veracruz,
Mexico

Rosendo Carrasco Gutierrez
Professor, Benemerita Universidad
Autonoma de Puebla, Facultad de
Estomatología, Puebla, Puebla,
Mexico

Rosario Jimenez Flores
Professor, Benemerita Universidad
Autonoma de Puebla, Facultad de
Estomatología, Puebla, Puebla,
Mexico

Diana Laura Saldaña Avalos
Dentistry Student, Universidad
Autonoma de Nuevo Leon,
Facultad de Odontología,
Monterrey, Nuevo Leon, 64460
ZIP, Mexico

Juan Manuel Solis Soto
Professor, Universidad Autonoma
de Nuevo Leon, Facultad de
Odontología, Monterrey, Nuevo
Leon, 64460 ZIP, Mexico

Corresponding Author:

Adrián Marcelo Villarreal García
Master's in Sciences Student,
Universidad Autonoma de
Nuevo Leon, Facultad de
Odontología, Monterrey, Nuevo
Leon, 64460 ZIP, Mexico

Veillonella parvula: An update from a stomatological point of view

Adrián Marcelo Villarreal García, Fanny Lopez Martinez, Laura Roesch Ramos, Evelyn Guadalupe Torres Capetillo, Rosendo Carrasco Gutierrez, Rosario Jimenez Flores, Diana Laura Saldaña Avalos and Juan Manuel Solis Soto

DOI: <https://doi.org/10.22271/oral.2022.v8.i4c.1641>

Abstract

Introduction: *Veillonella* species found in oral cavity are among the most abundant members of the oral flora, they are involved in bone and joint infections.

Objective: To analyze the literature on *V. parvula*, particularly bacteremia, treatment, colonization of dental implants, and symbiosis with other microorganisms.

Methodology: A review of the literature was carried out in the databases PubMed, Google Scholar and SCOPUS with the words "*Veillonella parvula* and Treatment and Clinical Advancements and biological markers, and dental implants".

Results: *Veillonella parvula* has different clinical manifestations and usually presents in a variety of systemic diseases. Among its treatment is the use of antibiotics, surgical lavage and photodynamic therapy. This bacterium is a very effective colonizer of dental implants of different composition. It is related to several oral microorganisms, including *Streptococcus*, *Fusobacterium nucleatum* and *Aggregatibacter actinomycetemcomitans*.

Conclusion: It is important to identify *V. parvula* for proper treatment, as it exacerbates various oral diseases by making symbiosis with other oral pathogens.

Keywords: *Veillonella parvula*", "bacteremia", "colonization", "symbiosis", "treatment

1. Introduction

Veillonella parvula is a bacterium that can cause microbial spondylarthritis; it is a poorly studied bacterium because it is so rare [1]. *Veillonella parvula* is a gram-negative anaerobic coccus that is rarely involved in bone and joint infections. It is a commensal and opportunistic pathogen that inhabits complex human microbial communities, including the gut and dental plaque microbiota. It has also been observed to have anaerobic anthracene biodegradation activity in non-smoking patients, and because of this the microbiome of the oral cavity is not affected [2, 3, 4]. This bacterium is a high colonizer in endodontic infections [5]. *Veillonella parvula* has been found not to colonize dental implants as successfully [6] and may cause preorbital cellulitis [7]. *Veillonella parvula* may be involved in caries development through interactions with *Streptococcus mutans* [8], is closely linked to hepatic encephalopathy [9] and has also been found to promote intestinal inflammation [10].

Veillonella parvula has a natural competence ability that is highly influenced by the composition of the growth medium and is useful for genetic manipulation of the bacterium [11]. There is no adequate review of *Veillonella parvula* in the literature from a stomatological point of view, therefore, the objective is to review the importance of this bacterium on dental field, particularly; bacteremia, treatment, dental implant and symbiosis with other microorganisms.

2. Materials and Methods

Articles on the subject published through the PubMed, SCOPUS and Google Scholar databases were analyzed, with emphasis on the last 5 years. The quality of the articles was evaluated using guidelines, i.e., identification, review, choice and inclusion.

The quality of the reviews was assessed using the measurement tool for evaluating systematic reviews [12]. The search was performed using Boolean logical operators AND, OR and NOT; with the keywords: "*Veillonella parvula*", "bacteremia", "treatment", "dental implant", "microbiota". The keywords were used individually, as well as each of them related to each other.

3. Results and Discussion

3.1 Bacteremia Due to *Veillonella parvula*

It has been documented that *Veillonella parvula* can cause spondylitis [1], osteomyelitis [3], and epidural abscesses in immunocompromised patients [13]. This bacterium has been seen in children with pneumonia [14] and acromioclavicular septic arthritis [15], reduces the activity of bronchial epithelia in obstructive pulmonary disease [16, 17] and produces nasal sepsis [18]. It has also been found at a high level in patients with COVID 19 [19], as well as with early childhood caries [20], lung cancer [21], nasopharyngeal carcinoma [22], and epithelial squamous cell carcinoma [23]. It has effects in patients with cell death in oral epithelium [24] and with severe alcoholic hepatitis [25], while it was increased in patients using proton pump inhibitors as treatment [26]. This bacterium is also elevated in the same way in patients with Sjorgen's syndrome [27] and with nephropathies [28].

Several authors conclude that *Veillonella parvula* bacteremia can cause different clinical manifestations and symptomatology in patients, such as spondylitis, arthritis, reduced bronchial activity and nasal sepsis in patients with different ages and comorbidities. Likewise, this bacterium seems to be increased in different systemic diseases such as COVID-19, cancers, etc.

3.2 Treatment

Veillonella parvula is generally considered a contaminating bacterium associated with polymicrobial infection. Antibiotics recommended in the therapy of infections caused by *Veillonella* species include penicillins, metronidazole, cephalosporins, aminoglycosides, imipenem, clindamycin, doxycycline, erythromycin, and chloramphenicol [29]. The use of ceftriaxone together with amoxicillin with clavulanic acid has also been shown to be effective in patients with endocarditis with bilateral abscess and osteomyelitis in a 6-month recovery [30]. However, in another investigation in anaerobic bacteria, including *Veillonella parvula*, greater pharmacological tolerance to amoxicillin and clindamycin was found, presumably due to metabolic changes of the bacteria residing within the biofilm [31]. In addition, in another study, when treating fibrous cystitis with antibiotics, *Veillonella parvula* and other bacteria were not affected [32]. This bacterium is resistant to herbal treatments, such as *Armoracia rusticana* and *Capuchina* [33].

In particular cases, such as *Veillonella parvula* bacteremia making a nasal abscess, drainage may help to relieve symptoms [18]. Surgical lavage has been shown to be effective in patients with sepsis of the acromioclavicular joint infected by *Veillonella parvula* [15]. Photodynamic therapy has been shown to be effective in the treatment of this bacterium [34] and a toothpaste with amorphous calcium phosphopeptide-phosphate in the mouth has also shown effectiveness [35].

Veillonella parvula shows different bacterial resistances and has different types of treatment, such as drainage and surgical lavage, the use of pastes and photodynamic therapy is equally effective. The use of different antibiotics in hospital use has shown efficacy on certain occasions and more research is

required to determine the best treatment.

3.3 Colonization of dental implants

Veillonella parvula is an early colonizer of dental implants in adult patients. Implants with moderately rough surfaces accumulated more bacterial biomass and significantly higher numbers of pathogenic bacteria [36]. Sloped implants had significantly higher mean probing depth and plaque index compared to axial implants [37]. Biofilm control of dental implants is hampered by their design and can cause oral and systemic problems, mainly in immunocompromised patients such as the elderly. Knowledge about the microbiota reinforces awareness of the need for periodic maintenance of professional cleaning [38]. In a study of 46 young people, they found that this bacterium is a colonizer on implants in young patients [39]. *Veillonella parvula* together with *Fusobacterium nucleatum* colonize implants on the first day of placement [40]. This bacterium has adherence to sterile titanium, hydroxyapatite and zirconium implants [41].

To reduce colonization on dental implants, titanium disc antibiotics have been used with implant solutions in combination with 0.12% chlorhexidine brushing and 0.05% cetylpyridinium chloride mouthwash, measured with polymerase, and showed a reduction of bacteria at 48 and 72 hours [42]. Probiotics have also been shown to be effective in drastically reducing biofilm formation by secondary colonizers that act as bridges, thus inhibiting biofilm formation on the titanium surface [43]. Peptide-based coatings have been shown to reduce the colonization of *Veillonella parvula* and other bacteria by 25% [44].

This bacterium is a colonizer of dental implants of different composition, there have been investigations on how to avoid its colonization and all have shown positive results, mainly with antibiotics and chlorhexidine.

3.4 Symbiosis with Other microorganisms

The *Veillonella* family, including *Veillonella parvula*, can form a biofilm with different *Streptococcus* families such as *Streptococcus mutans*, *Streptococcus gordonii* and *Streptococcus salivarius* [45]. *Veillonella* acts as a bridge to different bacteria such as *Streptococcus gordonii* and *Fusobacterium nucleatum*, producing nutrients for the survival and growth of periodontal pathogens [46]. In one study, *Veillonella parvula*, *Fusobacterium nucleatum* and *Aggregatibacter actinomycetemcomitans* were found to have interactions with each other, making a mutualism for the progression of periodontal disease [47]. Another investigation showed that there is coaggregation of *Veillonella parvula* with *Streptococcus gordonii* in saliva [48]. Likewise, several studies demonstrated that the *Veillonella* family coaggregates with the *Streptococcus* family [49]. As well as *Veillonella parvula* was found to interact with *Streptococcus mutans* in helping with dental caries [8]. In one study, it was shown that between 4 and 8 hours after plaque development, dominant strains of *Veillonella* change in their phenotypic characteristics (Coaggregation and antibody reactivity), as well as in their genotypic characteristics (16S RNA gene sequences, as well as strain-level fingerprinting patterns). This succession is coordinated with the development of mixed-species bacterial colonies. Changes in community structure can occur very rapidly in the development of natural biofilms, and it is suggested that this process may influence evolution within this ecosystem [50].

Veillonella parvula interacts with other bacteria such as *Streptococcus*, *Fusobacterium nucleatum* and

Aggregatibacter actinomycetemcomitans to form biofilm communities and thus, manifest oral pathologies such as caries and periodontal disease.

4. Conclusions

Veillonella parvula is a poorly studied bacterium that can manifest itself differently in symptomatology. Its treatment is mainly with antibiotics, although it is still being studied to determine the most effective way to eliminate it. This bacterium has been increased with different diseases and has been shown to colonize dental implants of different composition. It interacts with *Streptococcus*, *Fusobacterium* and *Aggregatibacter*, forming a biofilm in the oral cavity creating a variety of pathologies.

Author's Contribution

Not available

Conflict of Interest

Not available

Financial Support

Not available

4. References

- Ziga M, Gianoli D, Waldeck F, Dennler C, Schlichtherle R, Forster T, *et al.* Spondylodiscitis due to anaerobic bacteria *Veillonella parvula*: Case report and literature review. *Surg Neurol Int.* 2021 Sep 30;12:496.
- Béchon N, Jiménez-Fernández A, Witwinowski J, Bierque E, Taib N, Cokelaer T, *et al.* Autotransporters Drive Biofilm Formation and Autoaggregation in the Diderm Firmicutes *Veillonella parvula*. *J Bacteriol.* 2020 Oct 8;202(21):e00461-20.
- Gouze H, Noussair L, Padovano I, Salomon E, de Laroche M, Duran C, *et al.* *Veillonella parvula* spondylodiscitis. *Med Mal Infect.* 2019 Feb;49(1):54-58.
- Moussa HA, Wasfi R, Abdeltawab NF, Megahed SA. High Counts and Anthracene Degradation Ability of *Streptococcus mutans* and *Veillonella parvula* Isolated From the Oral Cavity of Cigarette Smokers and Non-smokers. *Front Microbiol.* 2021 Jun 28;12:661509.
- Pourhajibagher M, Ghorbanzadeh R, Bahador A. Culture-dependent approaches to explore the prevalence of root canal pathogens from endodontic infections. *Braz Oral Res.* 2017 Dec 18;31:e108.
- Saha S, Meenawat A, Sahu C, Srivastava V, Yadav S, Kumar V. Bacterial Identification and Monitoring Around Two-Piece Dental Implants by Matrix-Assisted Laser Desorption Ionization Time of Flight Mass Spectrometry (MALDI-TOF MS). *J Lab Physicians.* 2020 Mar;12(1):49-55.
- Wellens L, Casteels I, Huygens M. *Veillonella parvula* periorbital cellulitis: an unusual pathogen causing a common clinical sign. *GMS Ophthalmol Cases.* 2019 May 29;9:Doc17.
- Liu S, Chen M, Wang Y, Zhou X, Peng X, Ren B, *et al.* Effect of *Veillonella parvula* on the physiological activity of *Streptococcus mutans*. *Arch Oral Biol.* 2020 Jan;109:104578.
- Sung CM, Lin YF, Chen KF, Ke HM, Huang HY, Gong YN, *et al.* Predicting Clinical Outcomes of Cirrhosis Patients With Hepatic Encephalopathy From the Fecal Microbiome. *Cell Mol Gastroenterol Hepatol.* 2019;8(2):301-318.e2.
- Zhan Z, Liu W, Pan L, Bao Y, Yan Z, Hong L. Overabundance of *Veillonella parvula* promotes intestinal inflammation by activating macrophages via LPS-TLR4 pathway. *Cell Death Discov.* 2022 May 6;8(1):251.
- Knapp S, Brodal C, Peterson J, Qi F, Kreth J, Merritt J. Natural Competence Is Common among Clinical Isolates of *Veillonella parvula* and Is Useful for Genetic Manipulation of This Key Member of the Oral Microbiome. *Front Cell Infect Microbiol.* 2017 Apr 20;7:139.
- Shea BJ, Reeves BC, Wells G, Thuku M, Hamel C, Moran J, *et al.* AMSTAR 2: a critical appraisal tool for systematic reviews that include randomised or non-randomised studies of healthcare interventions, or both. *BMJ.* 2017;358:j4008.
- Kurihara M, Tamaki I, Tokuda Y. Epidural abscess and spondylitis caused by *Veillonella parvula* in a man on hemodialysis. *Clin Case Rep.* 2021 Oct 8;9(10):e04660.
- Pettigrew MM, Kwon J, Gent JF, Kong Y, Wade M, Williams DJ, *et al.* Antibacterial Resistance Leadership Group. Comparison of the Respiratory Resistomes and Microbiota in Children Receiving Short versus Standard Course Treatment for Community-Acquired Pneumonia. *mBio.* 2022 Apr 26;13(2):e0019522.
- Prod'homme M, Pfander G, Pavese P, Cech A, Abouelnaga I, Helfer L. Acromioclavicular Septic Arthritis Caused by *Veillonella parvula*. *Case Rep Orthop.* 2019 Nov 27;2019:7106252.
- Ke L, Chen L, Yaling Y, Can G, Jun L, Chuan Z. Investigation on the Pathological Mechanism of Frequent Exacerbators With Chronic Obstructive Pulmonary Disease Based on the Characteristics of Respiratory Flora. *Front Med (Lausanne).* 2022 Jan 20;8:816802.
- Chen YC, Ko PH, Yang CJ, Chen YC, Lay CJ, Tsai CC, *et al.* Epidural abscess caused by *Veillonella parvula*: Case report and review of the literature. *J Microbiol Immunol Infect.* 2016 Oct;49(5):804-808.
- Hyo Y, Fukushima H, Harada T, Hara H. Nasal septal abscess caused by anaerobic bacteria of oral flora. *Auris Nasus Larynx.* 2019 Feb;46(1):147-150.
- Ma S, Zhang F, Zhou F, Li H, Ge W, Gan R, *et al.* Metagenomic analysis reveals oropharyngeal microbiota alterations in patients with COVID-19. *Signal Transduct Target Ther.* 2021 May 13;6(1):191.
- Grier A, Myers JA, O'Connor TG, Quivey RG, Gill SR, Kopycka-Kedzierawski DT. Oral Microbiota Composition Predicts Early Childhood Caries Onset. *J Dent Res.* 2021 Jun;100(6):599-607.
- Tsay JJ, Wu BG, Sulaiman I, Gershner K, Schluger R, Li Y, *et al.* Lower Airway Dysbiosis Affects Lung Cancer Progression. *Cancer Discov.* 2021 Feb;11(2):293-307.
- Jiang H, Li J, Zhang B, Huang R, Zhang J, Chen Z, Shang X, Li X, Nie X. Intestinal Flora Disruption and Novel Biomarkers Associated With Nasopharyngeal Carcinoma. *Front Oncol.* 2019 Dec 6;9:1346.
- Rai AK, Panda M, Das AK, Rahman T, Das R, Das K, *et al.* Dysbiosis of salivary microbiome and cytokines influence oral squamous cell carcinoma through inflammation. *Arch Microbiol.* 2021 Jan;203(1):137-152.
- White T, Alimova Y, Alves VTE, Emecen-Huja P, Al-Sabbagh M, Villasante A, *et al.* Oral commensal bacteria differentially modulate epithelial cell death. *Arch Oral Biol.* 2020 Dec;120:104926.
- Kim SS, Eun JW, Cho HJ, Song DS, Kim CW, Kim YS, *et al.* Microbiome as a potential diagnostic and predictive biomarker in severe alcoholic hepatitis. *Aliment Pharmacol Ther.* 2021 Feb;53(4):540-551.
- Horvath A, Rainer F, Bashir M, Leber B, Schmerboeck B, Klymiuk I, *et al.* Biomarkers for oralization during

- long-term proton pump inhibitor therapy predict survival in cirrhosis. *Sci Rep.* 2019 Aug 19;9(1):12000.
27. Singh M, Teles F, Uzel NG, Papas A. Characterizing Microbiota from Sjögren's Syndrome Patients. *JDR Clin Trans Res.* 2021 Jul;6(3):324-332.
 28. Li X, Wang L, Ma S, Lin S, Wang C, Wang H. Combination of *Oxalobacter Formigenes* and *Veillonella parvula* in Gastrointestinal Microbiota Related to Bile-Acid Metabolism as a Biomarker for Hypertensive Nephropathy. *Int J Hypertens.* 2022 May 17;2022:5999530.
 29. Li J, Chen P, Li J, Gao X, Chen X, Chen J. A new treatment of sepsis caused by *Veillonella parvula*: A case report and literature review. *J Clin Pharm Ther.* 2017 Oct;42(5):649-652.
 30. Richards T, Stephen J, Lui CL. Severe disseminated *Veillonella parvula* infection including endocarditis, bilateral psoas abscess, discitis, and osteomyelitis but sparing spinal and hip prostheses: a case report. *J Med Case Rep.* 2022 Apr 20;16(1):157.
 31. Wuersching SN, Huth KC, Hickel R, Kollmuss M. Targeting antibiotic tolerance in anaerobic biofilms associated with oral diseases: Human antimicrobial peptides LL-37 and lactoferricin enhance the antibiotic efficacy of amoxicillin, clindamycin and metronidazole. *Anaerobe.* 2021 Oct;71:102439.
 32. Vandeplassche E, Sass A, Ostyn L, Burmølle M, Kragh KN, Bjarnsholt T, Coenye T, Crabbé A. Antibiotic susceptibility of cystic fibrosis lung microbiome members in a multispecies biofilm. *Biofilm.* 2020 Jun 13;2:100031.
 33. Eichel V, Schüller A, Biehler K, Al-Ahmad A, Frank U. Antimicrobial effects of mustard oil-containing plants against oral pathogens: an *in vitro* study. *BMC Complement Med Ther.* 2020 May 24;20(1):156.
 34. Kang SM, Jung HI, Kim BI. Susceptibility of oral bacteria to antibacterial photodynamic therapy. *J Oral Microbiol.* 2019 Jul 19;11(1):1644111.
 35. Philip N, Leishman SJ, Bandara HMHN, Healey DL, Walsh LJ. Randomized Controlled Study to Evaluate Microbial Ecological Effects of CPP-ACP and Cranberry on Dental Plaque. *JDR Clin Trans Res.* 2020 Apr;5(2):118-126.
 36. Bermejo P, Sánchez MC, Llama-Palacios A, Figuero E, Herrera D, Sanz Alonso M. Biofilm formation on dental implants with different surface micro-topography: An *in vitro* study. *Clin Oral Implants Res.* 2019 Aug;30(8):725-734.
 37. Narvaja A, Shibli JA, Coppede A, Giro G, Feres M, Favari M. Microbiologic Analysis of Immediately Loaded Full-Arch Implant-Retained Prosthesis Protocol After 2 Years of Loading: A Retrospective Study. *Int J Oral Maxillofac Implants.* 2018 Nov/Dec;33(6):1339-1344.
 38. Matthes de Freitas Pontes K, Fontenelle ISO, Nascimento CD, Oliveira VC, Albuquerque Garcia B, Silva PGB, *et al.* Clinical study of the biofilm of implant-supported complete dentures in healthy patients. *Gerodontology.* 2022 Jun;39(2):148-160.
 39. Dierens M, Vandeweghe S, Kisch J, Persson GR, Cosyn J, De Bruyn H. Long-term follow-up of turned single implants placed in periodontally healthy patients after 16 to 22 years: microbiologic outcome. *J Periodontol.* 2013 Jul;84(7):880-94.
 40. Siddiqui DA, Fidai AB, Natarajan SG, Rodrigues DC. Succession of oral bacterial colonizers on dental implant materials: An *in vitro* biofilm model. *Dent Mater.* 2022 Feb;38(2):384-396.
 41. Sánchez MC, Llama-Palacios A, Fernández E, Figuero E, Marín MJ, León R, *et al.* An *in vitro* biofilm model associated to dental implants: structural and quantitative analysis of *in vitro* biofilm formation on different dental implant surfaces. *Dent Mater.* 2014 Oct;30(10):1161-71.
 42. Virto L, Simões-Martins D, Sánchez MC, Encinas A, Sanz M, Herrera D. Antimicrobial effects of a new brushing solution concept on a multispecies *in vitro* biofilm model growing on titanium surfaces. *Clin Oral Implants Res.* 2022 Feb;33(2):209-220.
 43. Kang MS, Park GY. *In vitro* Evaluation of the Effect of Oral Probiotic *Weissella cibaria* on the Formation of Multi-Species Oral Biofilms on Dental Implant Surfaces. *Microorganisms.* 2021 Nov 30;9(12):2482.
 44. Fang D, Yuran S, Reches M, Catunda R, Levin L, Febbraio M. A peptide coating preventing the attachment of *Porphyromonas gingivalis* on the surfaces of dental implants. *J Periodontal Res.* 2020 Aug;55(4):503-510.
 45. Mashima I, Nakazawa F. The influence of oral *Veillonella* species on biofilms formed by *Streptococcus* species. *Anaerobe.* 2014 Aug;28:54-61.
 46. Zhou P, Li X, Huang IH, Qi F. *Veillonella* Catalase Protects the Growth of *Fusobacterium nucleatum* in Microaerophilic and *Streptococcus gordonii*-Resident Environments. *Appl Environ Microbiol.* 2017 Sep 15;83(19):e01079-17.
 47. Periasamy S, Kolenbrander PE. *Aggregatibacter actinomycetemcomitans* builds mutualistic biofilm communities with *Fusobacterium nucleatum* and *Veillonella* species in saliva. *Infect Immun.* 2009 Sep;77(9):3542-51.
 48. Mutha NVR, Mohammed WK, Krasnogor N, Tan GYA, Wee WY, Li Y, *et al.* Transcriptional profiling of coaggregation interactions between *Streptococcus gordonii* and *Veillonella parvula* by Dual RNA-Seq. *Sci Rep.* 2019 May 21;9(1):7664.
 49. Chalmers NI, Palmer RJ Jr, Cisar JO, Kolenbrander PE. Characterization of a *Streptococcus* sp.-*Veillonella* sp. community micromanipulated from dental plaque. *J Bacteriol.* 2008 Dec;190(24):8145-54.
 50. Palmer RJ Jr, Diaz PI, Kolenbrander PE. Rapid succession within the *Veillonella* population of a developing human oral biofilm *in situ*. *J Bacteriol.* 2006 Jun;188(11):4117-24.

How to Cite This Article

MVG Adrián, LM Fanny, RR Laura, GTC Evelyn, CG Rosendo, JF Rosario, LSA Diana, MSS Juan. *Veillonella parvula*: An update from a stomatological point of view. *International Journal of Applied Dental Sciences.* 2022;8(4):169-172.

Creative Commons (CC) License

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 4.0 International (CC BY-NC-SA 4.0) License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.